6-Gingerol, a functional polyphenol of ginger, reduces pulmonary fibrosis by activating Sirtuin1

Li Liua, Nan Yub, Wei Lengb, Yun Lua, Xinxín Xia, Hui Yuanb*

aThe Affiliation Hospital of Chengdu University of Traditional Chinese Medicine, Chengdu, China
bThe Affiliated Hospital of Shaanxi University of Traditional Chinese Medicine, Xianyang, China
cThe First Affiliated Hospital of Xi’an Jiaotong University, Xi’an, China

Received 2 November 2021; Accepted 7 December 2021
Available online 1 March 2022

KEYWORDS
6-gingerol;
SIRT1;
pulmonary fibrosis;
MRC-5 cells;
生姜

Abstract
Pulmonary fibrosis in general is the final common outcome of various interstitial lung diseases. In recent years, the incidence of pulmonary fibrosis has been rising with poor prognosis. 6-gingerol is deemed as a functional polyphenol of ginger. The aim of the present study was to investigate the effect of 6-gingerol on pulmonary fibrosis. Mice were randomly divided into four groups: control, bleomycin, bleomycin + 6-gingerol 100 mg/kg, bleomycin + 6-gingerol 250 mg/kg, and the survival rates of the groups were recorded. Pathological and fibrotic changes in the lungs were identified by H&E and Masson staining, respectively. The levels of hydroxyproline and protein deposited in lung tissues were then, respectively, determined by colorimetry and western blotting. Subsequently, the proportion of cells and inflammatory factors in the alveolar lavage fluid were estimated. Following the identification of the possibility of Sirtuin1 (SIRT1) in the pharmacological mechanism through molecular docking and western blotting, human embryonic lung fibroblasts MRC-5 were treated with TGF-β1 and SIRT1 inhibitor to study the role of SIRT1 in the regulatory effect of 6-gingerol. From the results, 6-gingerol was found to increase the survival rate of mice and reduce lung pathology and fibrosis in mice. And, it significantly reduced the levels of hydroxyproline and the proteins deposited in lung tissues. Moreover, the number of neutrophils, basophils, monocytes, and the levels of inflammatory factors in the alveolar lavage fluid were also reduced. SIRT1 inhibitor blocked the function of 6-gingerol to inhibit fibrosis. To sum up, 6-gingerol relieves pulmonary fibrosis via activating SIRT1. This finding expands the pharmacological effect of 6-gingerol, and it is expected to advance the development of treatments for pulmonary fibrosis.

© 2022 Codon Publications. Published by Codon Publications.
Introduction

The lung is composed of bronchi, alveoli, blood vessels, nerves, and lymph and connective tissue. Among them, alveoli and bronchi belong to the lung parenchyma, and the rest are parts of the pulmonary interstitium, which is distributed between the parenchyma and plays a role in connection, fixation, and nutrition. Viral infection, air pollution, smoking, and immune system abnormalities are all potential risk factors for interstitial lung diseases. Due to the unpredictability of interstitial lung diseases, daily attention should be paid to reducing the erosion of pathogenic factors. Moreover, pulmonary fibrosis is often the final common outcome of various interstitial lung diseases. In recent years, the incidence of pulmonary fibrosis has been on the rise, and its poor prognosis is manifested by the low average survival time of less than 5 years after diagnosis. Delaying the progress of pulmonary fibrosis and reducing its incidence are major needs for disease prevention and treatment due to the lack of effective radical treatment. It is well documented that the accumulation of inflammatory factors, the collapse of blood vessels, the increase of myofibroblasts, the deposition of extracellular matrix, and the final fibrosis are pivotal factors contributing to the pathogenesis of pulmonary fibrosis. Therefore, seeking effective treatment targets and lead compounds have great clinical significance for reducing the incidence of pulmonary fibrosis.

Ginger is a kind of rhizome-type spice seasoning that is widely cultivated worldwide. It is extensively applied in food processing and food preparation. Notably, people in numerous countries are already accustomed to treating flu, typhoid, and cough by making ginger tea. Contemporary scientific researchers have confirmed that the component gingerol accounts for the pharmacological properties of ginger. The gingerol isolated from ginger is a mixture of multiple substances. According to the chemical structure, gingerol is divided into two categories: single aromatic ring type and double aromatic ring heptane type. The main difference between them is the aromatic ring substituent. There are currently more than 10 specific gingerols, all of which have similar properties and structures. Among them, 6-gingerol, the content of which accounts for more than 80% of the total gingerol, is the most important component. Multiple studies have demonstrated that 6-gingerol may be capable of ameliorating hypoglycemia and exerting anti-hyperlipidemia, antioxidant, and anti-inflammatory roles. Furthermore, 6-gingerol has been found to reduce isoproterenol-induced myocardial fibrosis. Therefore, the hypothesis that 6-gingerol could also reduce pulmonary fibrosis was proposed in this work.

In the present study, for the purpose of studying the effect of 6-gingerol on pulmonary fibrosis, mice and MRC-5 cells were separately induced with bleomycin (BLM) or TGF-B1 to establish pulmonary fibrosis models. The experimental protocol is shown in Figure 1. The findings of this research were expected to expand the pharmacological effect of 6-gingerol, and advance the development of treatments for pulmonary fibrosis.

Materials and Methods

Mice and in vivo model establishment

All animal experiments were carried out in strict compliance with the National Institutes of Health guidelines for the handling of laboratory animals and approved by the Ethics Committee of our Hospital. A total of 80 male C57BL/6 mice (aged 6-8 weeks; Cyagen Biosciences, Suzhou, China) were adaptively housed in an SPF environment with a 12-h light/dark cycle, relative humidity of 55%, and free access to food and water at 25°C for a week.

The mice were randomly divided into four groups: control, BLM, BLM + 6-gingerol 100 mg/kg, BLM + 6-gingerol 250 mg/kg. BLM group: for the induction of fibrosis, BLM (purity > 98.00%, GlpBio, United States) was dissolved in 50 μL sterile PBS, and mice were intratracheally administered (2 mg/kg body weight) with a single dose of BLM. This induction lasted 21 days, once a day. BLM + 6-gingerol group: in addition to being treated with BLM, mice were given 6-gingerol (100 or 250 mg/kg, purity > 98.00%, Yuanye Biotechnology, Shanghai, China) for 21 days, once a day. The control group was given normal saline gavage. When 21 days or the ethical endpoints were reached, the mice were euthanized using an overdose of sodium pentobarbital (100 mg/kg, intraperitoneal injection) followed by cervical dislocation.

Cell lines and in vitro model establishment

Human embryonic lung fibroblasts MRC-5 were purchased from Procell Life Science & Technology (Wuhan, China) and cultured in DMEM (Gibco, United States) supplemented with 10% fetal bovine serum (FBS, Invitrogen, CA, United States), 1% penicillin/streptomycin (Invitrogen) in 5% CO2 atmosphere at 37°C.

The cells were randomly divided into different groups: control, TGF-B1 (Abmole, Shanghai, China), TGF-B1 + 6-gingerol, TGF-B1 + 6-gingerol + EX-527 [Sirtuin1 (SIRT1) inhibitor, purity > 98.00%, GlpBio, United States]. To induce fibrosis, cells were exposed to 10 ng/mL TGF-B1 for 48 h. Other experimental groups were exposed to 10 ng/mL TGF-B1 and 6-gingerol (10 or 20 μM) or EX-527 (10 μM). Untreated cells were used as the control group.

Hematoxylin-eosin (H&E) staining

H&E staining was performed using H&E staining kit (#C0105, Beyotime Institute of Biotechnology, China) according to the manufacturer’s instructions. After the mice were sacrificed, the left lung tissues were collected and fixed in 4% paraformaldehyde at 4°C for 6 h, dehydrated with gradient alcohol, embedded in paraffin, and cut into 4-μm slices. The sections were deparaffinized in xylene, and rehydrated in gradient alcohols followed by being stained with hematoxylin solution for 5 min at room temperature. After the sections were dehydrated with distilled water and stained with eosin for 30 s at room temperature, they were dehydrated further with gradient alcohol and became transparent.
using xylene for 5 min. Physiological changes of the samples were observed under a light microscope (magnification 400×, Leica, Germany).

**Masson staining**

This assay was performed using Masson staining kit (#DC0032, Leagene, Anhui, China) according to the manufacturer’s instructions. Tissue sections were obtained as aforementioned. Following the staining of sections with Weigert’s iron hematoxylin for 5 min, they were differentiated with acidic ethanol for 10 s. Then, sections were treated with the aniline blue solution for 1 min, followed by ponceau red magenta dyeing solution for 5 min. Subsequently, sections were washed with the phosphomolybdic acid solution for 1 min and counterstained with the aniline blue dyeing solution for 1 min, the results were observed after dehydration under a light microscope (magnification 400×, Leica, Germany).

**Determination of hydroxyproline concentration**

This assay was performed using an assay kit (#A030-2-1, Jiancheng Bioengineering Institute, Nanjing, China) according to the manufacturer’s instructions. Lung tissue (50 mg) was taken in a tube, and 1 mL of hydrolysis solution was added to the tube. The tube was placed in boiling water for 20 min, and the PH value of the solution was adjusted to 6.0-6.8. After the sample was bathed in 60°C water for 15 min, centrifuged at 3500 r/min for 10 min, the supernatant was collected to measure the absorbance at 550 nm using a microplate reader (Dojindo, Rockville, United States).

**Enzyme-linked immunosorbent (ELISA) assay**

The necks of mice were dissected and the tracheae were exposed, lavage was performed with 0.8 mL PBS for three times, and the alveolar lavage fluid was collected as described previously. The collected lavage fluid was centrifuged at 3500 r/min and 4°C for 10 min. The expression levels of TNF-α (#88-7324) and IL-1β (#88-7013; both Thermo Fisher Scientific, Shanghai, China) in the supernatant were determined using commercial kits according to the manufacturer’s instructions. The absorbance at 450 nm was measured using a microplate reader (Dojindo).

**Western blotting analysis**

Cultured cells and lung tissue samples from experimental mice were lysed in RIPA lysis buffer (Beyotime Institute
of Biotechnology, China). After the determination of protein concentration with a BCA protein assay kit (#A045-3-2, Nanjing JianCheng Bioengineering Institute, Nanjing, China), 20 μg of protein per lane was separated by 10% SDS-PAGE. The separated proteins were transferred onto PVDF membranes, and the membranes were blocked with 5% skimmed milk at room temperature for 2 h. Subsequently, they were continued to incubate with primary antibodies against α-SMA (#ab5694, 1:200), collagen IV (#ab6586, 1:1000), SIRT1 (#ab189494, 1:1000), and GAPDH (#ab9485, 1:2500) at 4°C overnight, followed by HRP-conjugated anti-rabbit antibody (#ab97051; 1: 10,000) at 37°C for 1.5 h. All antibodies were acquired from Abcam. The membranes were developed with an ECL reagent (Millipore) at room temperature for 2 min. The blots were quantified using ImageJ software (v1.8; National Institutes of Health).

**Reverse transcription-quantitative PCR (RT-qPCR)**

Total RNA was extracted from cultured cells using TRIzol® reagent (Invitrogen). The extracted RNA was reverse transcribed into cDNA using a PrimeScript RT reagent kit (Takara, Japan) according to the manufacturer’s protocol. The mRNA levels were quantified using a QuantiTect SYBR Green PCR Kit (Qiagen) with the ABI7500 system (Applied Biosystems). The PCR amplification conditions were as follows: 95°C for 10 min and then 40 cycles of 95°C for 30 s, 64°C for 34 s, and 72°C for 30 s. The relative mRNA level was normalized to GAPDH and calculated using the 2^{ΔΔCq} method. The used primers for PCR are listed in Table 1.

**Cell counting kit-8 (CCK-8) assay**

Cell viability was determined using a CCK-8 assay. Cells (1 × 10^4 cells/well) were seeded in 96-well plates. After cultivation for 24 h, the normal medium was replaced with DMEM medium containing TGF-β1 (10 ng/mL), 6-gingerol (10 or 20 μM), or EX-527 (10 μM). Cells were continued to be incubated in a humidified incubator at 37°C for 24, 48, or 72 h. Afterwards, 10 μL of CCK-8 solution was added to each well for a further incubation for 2 h at 37°C. The optical density values were recorded at 450 nm using a microplate reader (Dojindo).

**Molecular docking**

Molecular docking was used to evaluate the binding force between 6-gingerol and SIRT1 protein. AutoDock v4.2 software (Scrpps Institute) was used in this study to perform molecular docking simulation. The three-dimensional (3D) structure of 6-gingerol was imported into the software and adjusted to display hydrogen atoms. The 3D structure of the protein was downloaded from the Protein Data Bank (PDB) website (https://www.rcsb.org/) and imported into the software. The water molecules and excess residues or ligands in the protein file were removed. After setting the docking position, the molecular docking was operated, and the docking energy value of the two was displayed after the operation was over.

**Statistical analysis**

Data were presented as means ± standard deviations (SD) of three experiments. One-way ANOVA with Tukey’s post-hoc test was used to compare differences among multiple groups. Statistical analysis was performed using GraphPad Prism 8.0 software, and the value of p < 0.05 was considered statistically significant.

**Results**

6-Gingerol improves the survival rate of bleomycin-induced mice and reduces fibrosis

The number of surviving mice in the four groups was recorded every day for a total of 21 days, and the survival rate was exhibited in the form of a line graph. The red line represented the survival rate of the BLM group, and blue and black lines represented the 6-gingerol treatment group at different concentrations. The daily survival rate of the 6-gingerol group was higher than that of the BLM group, and the high-concentration group was more effective in improving the survival of mice (Figure 2A). And, the changes of lung pathology in each group was observed by H&E staining. In the control group, the alveolar morphology was normal, the wall was slender, and there was no inflammatory cell infiltration in the lung interstitium. However, obvious pulmonary fibrosis changes were seen in the BLM group, manifested as sheet consolidation, with much inflammatory cell infiltration, fibroblast proliferation, and collagen deposition in the consolidation area. Simultaneously, less consolidation and fewer inflammatory cells were observed in the 6-gingerol 100 mg/kg treatment group than in the BLM group. More importantly, in the 6-gingerol 250 mg/kg treatment group, there was no pulmonary fibrosis changes of lung pathology in each group were observed by H&E staining. In the control group, the alveolar morphology was normal, the wall was slender, and there was no inflammatory cell infiltration in the lung interstitium. However, obvious pulmonary fibrosis changes were seen in the BLM group, manifested as sheet consolidation, with much inflammatory cell infiltration, fibroblast proliferation, and collagen deposition in the consolidation area. Simultaneously, less consolidation and fewer inflammatory cells were observed in the 6-gingerol 100 mg/kg treatment group than in the BLM group. More importantly, in the 6-gingerol 250 mg/kg treatment group, there was no...
Liu L et al.

The number of these cells in the 6-gingerol intervention groups was smaller than those in the BLM group, and the difference became more obvious in the high-dose group (p < 0.01; Figure 4A). Moreover, the expression levels of TNF-α and IL-1β in the alveolar lavage fluid were determined using ELISA assay. The results revealed that the levels of TNF-α and IL-1β in the BLM group were significantly upregulated compared with the control group (p < 0.001), while 6-gingerol intervention again cut down TNF-α and IL-1β levels (p < 0.01), and the finding was more apparent in the high-dose group (p < 0.001; Figure 4B).

SIRT1 is the probable target of 6-gingerol

Furthermore, the expression of SIRT1 in the lung tissues at the protein level was determined using western blotting. It was noticed that SIRT1 protein exhibited low expression in the BLM group relative to the control group (p < 0.001). Subsequently, the levels of hydroxyproline in the lung tissues were determined. The level of hydroxyproline in the BLM group was significantly higher than that of the control group, and hydroxyproline in the lung tissues of mice were significantly reduced by the administration of 6-gingerol (p < 0.001; Figure 3B). The levels of fibrotic deposition proteins were then determined by western blotting. The levels of α-SMA and collagen IV in the BLM group were significantly higher than those in the control group (p < 0.001), and 6-gingerol intervention could significantly reduce the expression levels of these two proteins (p < 0.01; Figure 3C).

6-Gingerol regulates the ratio of cells and inhibits the release of inflammatory factors in the alveolar lavage fluid

Elastase produced by neutrophils activates TGF-β and recruits inflammatory cells to the lungs, thereby promoting lung fibrosis.22 The number of basophils and monocytes is positively correlated with the progression of pulmonary fibrosis.24,25 From the experimental results, it could be seen that the number of neutrophils, basophils, and monocytes in the BLM group rose sharply (p < 0.001), while the number of these cells in the 6-gingerol intervention groups was smaller than those in the BLM group, and the difference became more obvious in the high-dose group (p < 0.01; Figure 4A). Moreover, the expression levels of TNF-α and IL-1β in the alveolar lavage fluid were determined using ELISA assay. The results revealed that the levels of TNF-α and IL-1β in the BLM group were significantly upregulated compared with the control group (p < 0.001), while 6-gingerol intervention again cut down TNF-α and IL-1β levels (p < 0.01), and the finding was more apparent in the high-dose group (p < 0.001; Figure 4B).

6-Gingerol reduces TGF-81-induced lung fibroblast proliferation and inflammatory and fibrotic matrix protein deposition by activating SIRT1

TGF-81 is a central mediator in the pathogenesis of pulmonary fibrosis.26 Through our investigation, SIRT1 protein...
Figure 3 6-Gingerol reduces lung fibrosis in bleomycin-induced mice. (A) Fibrotic changes in lung tissue were observed by masson staining. (B) The levels of hydroxyproline in the lung tissues were determined using colorimetry. (C) The levels of fibrotic deposition proteins were determined by western blotting. ***p < 0.001 vs. control; **p < 0.01, ###p < 0.001 vs. BLM.

level in TGF-β1-stimulated cells was downregulated compared with the control group (p < 0.001), while its protein level was enhanced in the 6-gingerol intervention group than that in the TGF-β1 group (p < 0.05; Figure 6A-B). Afterwards, cell viability was determined using CCK-8 assay. The results demonstrated that TGF-β1 promoted the viability of fibroblasts, while 6-gingerol exerted the opposite role in cell viability as TGF-β1. However, SITR1 inhibitor, EX-527, eased the inhibitory effect of 6-gingerol on cell viability (p < 0.01; Figure 6C). Besides, RT-qPCR analysis showed that the levels of inflammatory factors including TNF-α, IL-1β, and IL-6 were greatly increased in the TGF-β1 group (p < 0.001), and this effect was restored by 6-gingerol. Moreover, the addition of EX-527 could increase the levels of inflammatory factors again (p < 0.05; Figure 6D). Also, 6-gingerol significantly inhibited the overexpression of α-SMA and collagen IV induced by TGF-1β (p < 0.001), whereas this outcome was counteracted by EX-527 (p < 0.001; Figure 6E-F).

Discussion

Pulmonary fibrosis has the highest morbidity in population aged 40-50 years, and it tends to occur in men. The clinical manifestations are dyspnea, dry cough, and fatigue, accompanied by clubbing digits and cyanosis. The disease can be diagnosed based on clinical manifestations, imagological examination, and lung function tests. Currently, the treatment of patients with pulmonary fibrosis is basically not limited to one method, and a comprehensive treatment strategy is usually adopted, that is, the
Figure 4  6-Gingerol regulates the ratio of cells and inhibits the release of inflammatory factors in the alveolar lavage fluid. (A) The numbers of neutrophils, basophils, and monocytes in the alveolar lavage fluid of each group of mice were calculated. (B) The levels of TNF-α and IL-1β in the alveolar lavage fluid were determined using ELISA assay. ***p < 0.001 vs. control; ##p < 0.01, ###p < 0.001 vs. BLM.

Figure 5  SIRT1 is the probable target of 6-gingerol. (A) The expression levels of SIRT1 in the lung tissues were determined using western blotting. (B) Overall image and partial enlarged image of the results of molecular docking. ***p < 0.001 vs. control; ##p < 0.01 vs. BLM.
Figure 6  6-Gingerol reduces TGF-β1-induced lung fibroblast proliferation and inflammatory and fibrotic matrix protein deposition by activating SIRT1. (A) The expression levels of SIRT1 in the different groups of MRC-5 cells were determined using western blotting and (B) RT-qPCR. (C) Cell viability was determined using CCK-8 assay. (D) The expression levels of TNF-α, IL-1β, and IL-6 in the different groups of cells were determined using RT-qPCR. The expression levels of α-SMA and collagen IV in the different groups of cells were determined using (E) western blotting and (F) RT-qPCR. ***p < 0.001 vs. control; #p < 0.05, ##p < 0.01, ###p < 0.001 vs. TGF-β1; *p < 0.05, **p < 0.01, ***p < 0.001 vs. TGF-β1 + 6-gingerol.
combination of anti-pulmonary fibrosis drugs and nonpharmacological methods. Nonpharmacological methods include pulmonary rehabilitation (such as aerobic training, respiratory muscle training), oxygen therapy, invasive/noninvasive mechanical ventilation, and lung transplantation. However, the application of these methods is limited due to a list of disadvantages including high cost, low curative effect, insufficient transplantation donors, immune rejection, or poor patient compliance. Therefore, it is necessary to actively explore the pathogenesis of pulmonary fibrosis and develop novel intervention drugs.

The 6-gingerol used in this study is a natural medicinal component that exists in ginger. It was found to increase the survival rate of experimental mice and relieve pathological damage to the lungs in the mice models. Previous studies have revealed that 6-gingerol can reduce the damage of dextran sodium sulfate-induced ulcerative colitis, and relieve liver damage induced by sepsis. Moreover, 6-gingerol is believed to reduce the release of pro-inflammatory and pro-fibrotic factors during liver fibrosis. Although the target organs of the study are different, the finding in our study that 6-gingerol alleviated fibrosis coincides with the results of previous studies. Furthermore, higher concentration of 6-gingerol illustrated more effective inhibition of pulmonary fibrosis and improvement of survival rate of mice. This suggested that the effect of 6-gingerol on pulmonary fibrosis was concentration-dependent. Although, as a natural extract, 6-gingerol has superior safety in contrast to synthetic drugs, the dose of drug used is not negligible for the treatment. Whether elevating the experimental concentration of 6-gingerol will adversely affect the survival of mice requires further research.

In addition, we conducted a preliminary study on the mechanism of 6-gingerol on pulmonary fibrosis. Through computer simulation calculations, this paper found that 6-gingerol and SIRT1 had a stable binding through intermolecular forces. And, 6-gingerol markedly increased SIRT1 level in the lung tissue according to the western blot analysis. SIRT1 therein is a member of the Sirtuin family, which is a type of histone deacetylase that depends on NAD+. Interestingly, Conti et al. found that SIRT1 expression was downregulated in the lung tissue of patients with chronic obstructive pulmonary emphysema. Li et al. found that SIRT1 activation could alleviate lung injury caused by lipopolysaccharide. These findings indicated that the activation of SIRT1 was beneficial for normal lung function. Herein, 6-gingerol was found to inhibit lung fibroblast proliferation, reduce inflammation, and ensure fibrotic matrix protein deposition via activating SIRT1. The results of this article also illustrated the importance of SIRT1 in the lungs, and the thought of applying drugs to activate SIRT1 is an important and innovative approach to treat pulmonary fibrosis. Moreover, natural extract generally has little adverse effects to the human body, which is a significant advantage of its clinical application in the future.

Last but not least, SIRT1 has a variety of substrates, including transcription factors such as p53, p65, and Forkhead box O, nuclear receptors such as estrogen receptor and peroxisome proliferators-activated receptor; DNA repair protein components such as apurinic/apyrimidin endonuclease and NBS1; and intracellular signaling molecules such as signal transducer and activator of transcription. This article is limited to the discovery that SIRT1 is the target of 6-gingerol; however, what kind of substrate SIRT1 acts on to regulate pulmonary fibrosis will be the direction of our future research.

Conclusion

In conclusion, 6-gingerol was found to alleviate pulmonary fibrosis by activating SIRT1. This article revealed the role of 6-gingerol in suppressing pulmonary fibrosis for the first time. This discovery is expected to underline the importance of 6-gingerol and provide credible evidence that 6-gingerol may be used as a potential medicine.

Funding

This study was supported by The Hospital of Chengdu University of Traditional Chinese Medicine Scientific Research Capacity to enhance the “Hundred Talents Plan” (Li Liu, No. 20-Q06), The Discipline Innovation Team Project of Shaanxi University of Chinese Medicine (Nan Yu, No. 2019-QN09), Provincial Unit Chinese Medicine Development Fund-Provincial Famous Traditional Chinese Medicine Studio (Yun Lu, No. 2100601), Scientific Research Project of Affiliated Hospital of Shaanxi University of Chinese Medicine (Hui Yuan, No. 202QQ011) and National Natural Science Foundation of China Youth Fund (Xinxin Xia, No.81904073).

References


